

A Panel of Translational
Xenoreceptor, Cytochrome P450, Transporter
and Transplanted **Liver Humanized Mouse Models**
for Improved Prediction of Drug Responses in Man



Nico Scheer, 18th April 2013

Objectives



- **To provide a literature overview of genetically ADMET and liver humanized mouse models described to date**
- **To describe a systematic approach of generating genetically humanized ADMET mouse models and combining them into complex, multiple humanized genotypes**
- **To provide selected examples of the utility of these models**
- **To discuss the benefits & limitations of genetically ADMET and liver humanized mouse models**

Humanized Xenobiotic Receptor Mice



Receptor	Type of genetic modification	Promoter driving human gene expression	Human expression construct	Reference
PXR	KO + RT	Mouse albumin	cDNA	Xie et al., 2000
	RT	Mouse albumin	VP16-PXR cDNA	Xie et al., 2000
	KO + RT	Rat fatty acid-binding protein	cDNA	Zhou et al., 2006
	RT	Rat fatty acid-binding protein	VP16-PXR cDNA	Gong et al., 2006
	KO + RT	Human PXR	genomic	Ma et al., 2007
	KO + RT	Mouse transthyretin enhancer	cDNA	Lichti-Kaiser and Staudinger, 2008
	TR	Mouse Pxr	genomic/cDNA	Scheer et al., 2008
	TR	Mouse Pxr	genomic/cDNA	Scheer et al., 2010
	TR	Mouse PXR	partial cDNA	Igarashi et al., 2012
CAR	KO + RT	Mouse albumin	cDNA	Zhang et al., 2002
	TR	Mouse Car	genomic	Scheer et al., 2008
PPARα	KO + RT	Tetracyclin responsive regulatory element	cDNA	Cheung et al., 2004.
	KO + RT	Human PPAR α	genomic	Yang et al., 2008.
AHR	TR	Mouse Ahr	cDNA	Moriguchi et al., 2003
	KO + RT	Mouse transthyretin enhancer	cDNA	Flaveny et al., 2009

KO = knockout, RT = random transgenesis, TR = targeted replacement, VP16 = activation domain of herpes simplex.

Humanized DME & Transporter Mice



Human gene(s)	Features	Reference
CYP1A1 & 1A2	Random transgene of genomic human CYP1A1/1A2 sequence. Combined with knockout for mouse <i>Cyp1a1</i> or <i>Cyp1a2</i> .	(Cheung et al., 2005a; Jiang et al., 2005)
	Random transgene of genomic human CYP1A1/1A2 sequence. Combined with knockout for mouse <i>Cyp1a1/1a2</i> .	(Dragin et al., 2007)
CYP2A6	Random transgene of human CY2A6 cDNA expressed off the mouse transthyretin promoter/enhancer.	(Zhang et al., 2005)
CYP2A13 & 2B6/2F1	Random transgene of genomic human CYP2A13/2B6/2F1 sequence. Also combined with a mouse <i>Cyp2f2</i> knockout.	(Wei et al., 2012)
CYP2C9	Targeted replacement of the mouse <i>Cyp2c</i> locus with a genomic/cDNA hybrid of human CYP2C9. Human CYP2C9 is expressed off the mouse albumin promoter.	(Scheer et al., 2012b)
CYP2C18 & 2C19	Random transgene of genomic human CYP2C18/2C19 sequence.	(Lofgren et al., 2008)
CYP2D6	Random transgene of genomic human CYP2D6 sequence.	(Corchero et al., 2001)
	Targeted replacement of the mouse <i>Cyp2d</i> locus with a genomic human CYP2D6 sequence.	(Scheer et al., 2012c)
CYP2E1	Random transgene of human CYP2E1 cDNA expressed off the mouse albumin promoter.	(Morgan et al., 2002)
	Random transgene of genomic human CYP2E1 sequence. Combined with knockout for mouse <i>Cyp2e1</i> .	(Cheung et al., 2005b)
CYP3A4	Random transgene of genomic human CYP3A4 sequence.	(Granvil et al., 2003; Yu et al., 2005)
	Random transgene of human CYP3A4 cDNA expressed off the human ApoE promoter.	(van Herwaarden et al., 2005)
	Random transgene of human CYP3A4 cDNA expressed off the human ApoE promoter. Combined with knockout for mouse <i>Cyp3a</i> genes.	(van Herwaarden et al., 2007)
	Random transgene of human CYP3A4 cDNA expressed off the mouse villin promoter. Combined with knockout for mouse <i>Cyp3a</i> genes.	
	Random transgenes of human CYP3A4 cDNA expressed off the human ApoE and mouse villin promoter. Combined with knockout for mouse <i>Cyp3a</i> genes.	
CYP3A4 & 3A7	Random transgene of genomic human CYP3A4/CYP3A7 sequence.	(Cheung et al., 2006)
	Targeted replacement of the mouse <i>Cyp3a</i> locus with a genomic human CYP3A4/CYP3A7 sequence.	(Hasegawa et al., 2011a)
CYP3A4, 3A5, 3A7 & 3A43	Trans-chomosomal mice containing the human CYP3A cluster (CYP3A5 inactive). Combined with knockout for mouse <i>Cyp3a</i> genes.	(Kazuki et al., 2012)
CYP3A4 & 2D6	Random transgenes of genomic human CYP3A4 and CYP2D6 sequences.	(Felmlee et al., 2008)
CYP3A7	Random transgene of human CYP3A7 cDNA expressed off the mouse metallothionein-1 promoter.	(Li et al., 1996)
NAT2	Random transgene of human NAT2*4cDNA expressed off the rat probasin promoter.	(Leff et al., 1999)
	Random transgene of human NAT2*4 cDNA expressed off the mouse albumin promoter. Combined with knockout for mouse <i>Nat1/2</i> .	(Sugamori et al., 2011)
UGT1A	Random transgene of genomic human UGT1A sequence.	(Chen et al., 2005)
	Random transgenes of genomic human UGT1 ^{A1*1} or UGT1 ^{A1*28} sequences. Combined with knockout for mouse <i>Ugt1</i> .	(Fujiwara et al., 2010)
UGT2B7	Random transgene of genomic human UGT2B7 sequence.	(Yueh et al., 2011)
Human gene(s)	Features	Reference
MRP2	Targeted replacement of mouse <i>Mrp2</i> with a cDNA of human MRP2. Human MRP2 is expressed off the mouse <i>Mrp2</i> promoter.	(Scheer et al., 2012a)
OATP1B1	Random transgene of human OATP1B1 cDNA expressed off the human ApoE promoter.	(van de Steeg et al., 2009)
	Random transgene of human OATP1B1 cDNA expressed off the human ApoE promoter. Combined with knockout of mouse <i>Oatp1a/1b</i> genes.	(van de Steeg et al., 2012)
OATP1B3	Random transgene of human OATP1B3cDNA expressed off the human ApoE promoter. Combined with knockout of mouse <i>Oatp1a/1b</i> genes.	(van de Steeg et al., 2012)

Transplanted Humanized Liver Mice



Model	Immune deficiency	Induction of liver failure	Reference
uPA-Scid	SCID	Hepatic expression of urokinase-type plasminogen activator (uPA)	Mercer et al., 2001 Tateno et al., 2004
uPA-NOG	NOD/Shi-scid IL2Rg ^{null} (NOG)	Hepatic expression of urokinase-type plasminogen activator (uPA)	Suemizu et al., 2008
TK-NOG	NOD/Shi-scid IL2Rg ^{null} (NOG)	Hepatic HSVtk expression combined with Gancyclovir treatment	Hasegawa et al., 2011
FRG	Rag-2 ^{null} /IL2rg ^{null} (/NOD)	Deletion of fumarylacetoacetate hydrolase	Azuma et al., 2007

A Panel of Translational Xenoreceptor, DME, Transporter & Liver Humanized Mouse Models



Xenoreceptor Panel		Cytochrome P450 Panel		Transporter Panel	
Knockouts	Humanized	Knockouts	Humanized	Knockouts	Humanized
Ahr	AHR	Cyp1a1/1a2	CYP1A1/1A2	Bcrp	BCRP*
Car	CAR	Cyp2c	CYP2C9	Mdr1a, Mdr1a/1b	MDR1*
Ppara α		Cyp2d	CYP2D6	Bcrp/Mdr1a/1b	
Pxr	PXR	Cyp3a (BL/6)	CYP3A4/3A7	Mrp1	
Pxr/Car	PXR/CAR	Cyp3a (FVB)	Liver CYP3A4	Mrp2	MRP2
Pxr/Car/Ahr	PXR/CAR/AHR		Gut CYP3A4	Oatp1a/1b	OATP1B1
Pxr/Car/Ppara α	PXR/CAR/PPAR α		Liver/Gut CYP3A4		OATP1B3
Pxr/Car/Ahr/Ppara α	PXR/CAR/AHR/PPAR α	Cyp2c/Cyp2d/Cyp3a			OATP1B1/1B3
				Oct1/2	
				Oat1/3	OAT1/3

Phase 2 Panel	Composite models	Transplanted Liver Humanized Mice
Humanized	Humanized	
UGT1A1	PXR/CAR/CYP3A4/3A7	FRG™ (in Collaboration with Yecuris Inc.)
	PXR/CAR/CYP3A4/3A7/CYP2D6	
	PXR/CAR/CYP3A4/3A7/CYP2D6/CYP2C9	

*coming soon

A Panel of Translational Xenoreceptor, DME, Transporter & Liver Humanized Mouse Models

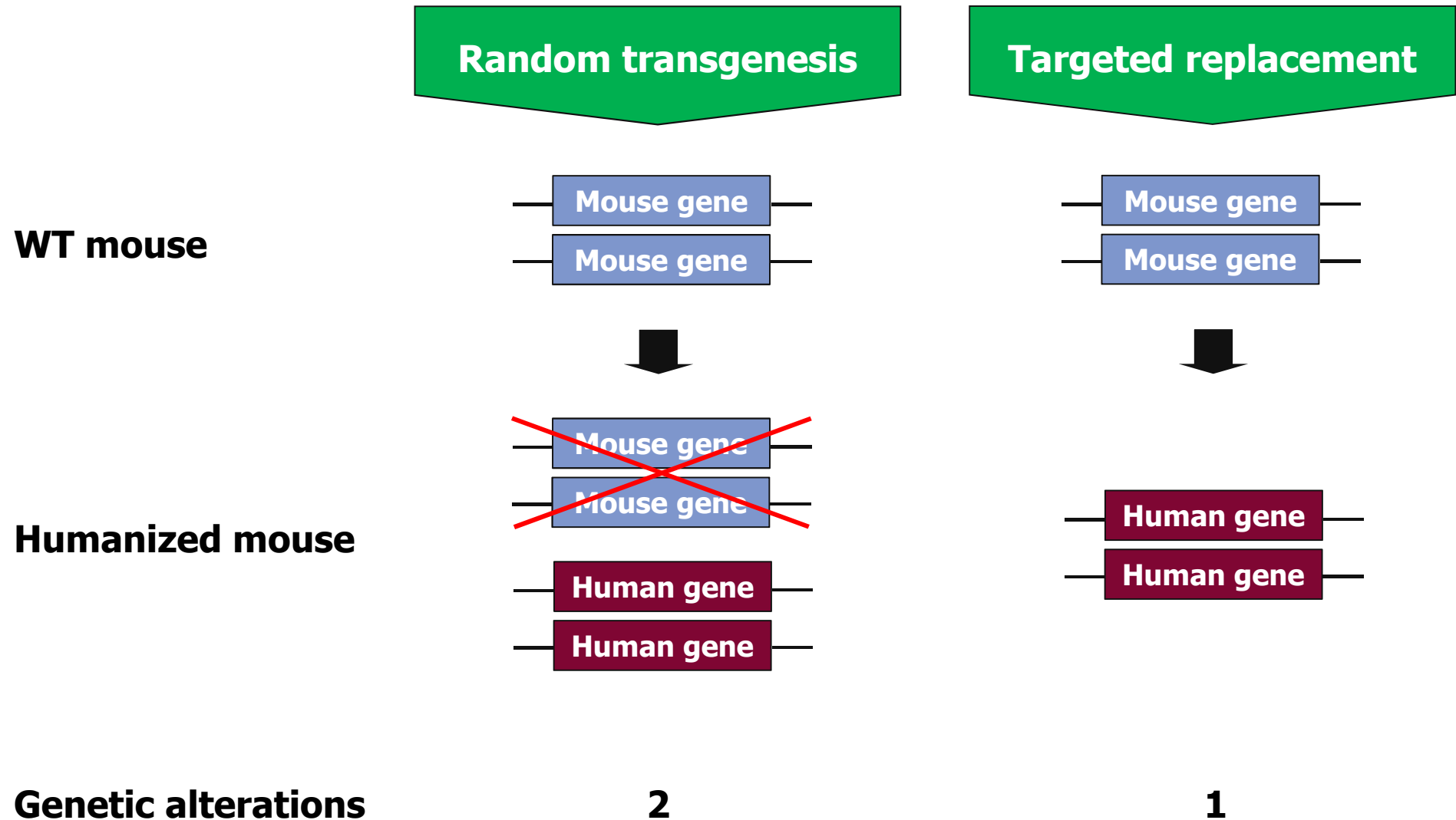


Xenoreceptor Panel		Cytochrome P450 Panel		Transporter Panel	
Knockouts	Humanized	Knockouts	Humanized	Knockouts	Humanized
Ahr	AHR	Cyp1a1/1a2	CYP1A1/1A2	Bcrp	BCRP*
Car	CAR	Cyp2c	CYP2C9	Mdr1a, Mdr1a/1b	MDR1*
Ppara α		Cyp2d	CYP2D6	Bcrp/Mdr1a/1b	
Pxr	PXR	Cyp3a (BL/6)	CYP3A4/3A7	Mrp1	
Pxr/Car	PXR/CAR	Cyp3a (FVB)	Liver CYP3A4	Mrp2	MRP2
Pxr/Car/Ahr	PXR/CAR/AHR		Gut CYP3A4	Oatp1a/1b	OATP1B1
Pxr/Car/Ppara α	PXR/CAR/PPAR α		Liver/Gut CYP3A4		OATP1B3
Pxr/Car/Ahr/Ppara α	PXR/CAR/AHR/PPAR α	Cyp2c/Cyp2d/Cyp3a			OATP1B1/1B3
				Oct1/2	
				Oat1/3	OAT1/3

Phase 2 Panel	Composite models	Transplanted Liver Humanized Mice
Humanized	Humanized	
UGT1A1	PXR/CAR/CYP3A4/3A7	FRG™ (in Collaboration with Yecuris Inc.)
	PXR/CAR/CYP3A4/3A7/CYP2D6	
	PXR/CAR/CYP3A4/3A7/CYP2D6/CYP2C9	

*coming soon

Approaches of generating genetically humanized mouse models (simplified)



A Panel of Translational Xenoreceptor, DME, Transporter & Liver Humanized Mouse Models

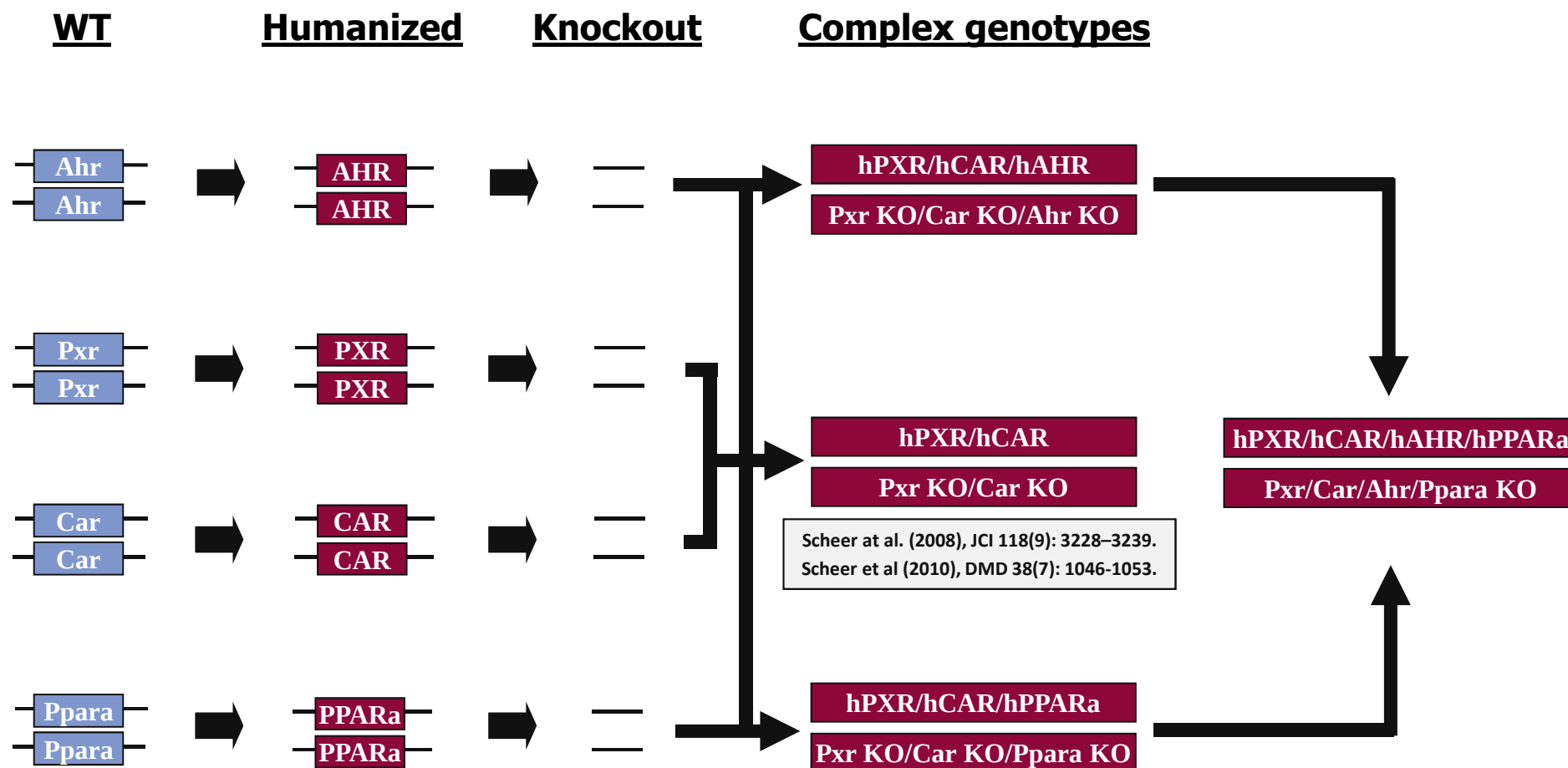


Xenoreceptor Panel		Cytochrome P450 Panel		Transporter Panel	
Knockouts	Humanized	Knockouts	Humanized	Knockouts	Humanized
Ahr	AHR	Cyp1a1/1a2	CYP1A1/1A2	Bcrp	BCRP*
Car	CAR	Cyp2c	CYP2C9	Mdr1a, Mdr1a/1b	MDR1*
Ppara α		Cyp2d	CYP2D6	Bcrp/Mdr1a/1b	
Pxr	PXR	Cyp3a (BL/6)	CYP3A4/3A7	Mrp1	
Pxr/Car	PXR/CAR	Cyp3a (FVB)	Liver CYP3A4	Mrp2	MRP2
Pxr/Car/Ahr	PXR/CAR/AHR		Gut CYP3A4	Oatp1a/1b	OATP1B1
Pxr/Car/Ppara α	PXR/CAR/PPAR α		Liver/Gut CYP3A4		OATP1B3
Pxr/Car/Ahr/Ppara α	PXR/CAR/AHR/PPAR α	Cyp2c/Cyp2d/Cyp3a			OATP1B1/1B3
				Oct1/2	
				Oat1/3	OAT1/3

Phase 2 Panel	Composite models	Transplanted Liver Humanized Mice
Humanized	Humanized	
UGT1A1	PXR/CAR/CYP3A4/3A7	FRG™ (in Collaboration with Yecuris Inc.)
	PXR/CAR/CYP3A4/3A7/CYP2D6	
	PXR/CAR/CYP3A4/3A7/CYP2D6/CYP2C9	

*coming soon

Multiple humanized xenoreceptor mice



Induction studies in xenoreceptor mice



PXR

[Rifampicin]
(mg/kg)

Cyp3a11 →

Cyp2b10 →

WT		hPXR	
0	10	0	10

[Dexamethasone]
(mg/kg)

Cyp3a11 →

WT			hPXR		
0	1	3	0	1	3

CAR

[CITCO]
(mg/kg)

Cyp2b10 →

WT			hCAR		
0	2,5	10	0	2,5	10

[TCPOBOP]
(mg/kg)

Cyp2b10 →

WT			hCAR		
0	0,3	1	0	0,3	1

AHR

	TCDD (µg/kg)			
	0	25	100	
C57BL/6 WT				Cyp1a1
hAHR				
Ahr KO				

A Panel of Translational Xenoreceptor, DME, Transporter & Liver Humanized Mouse Models

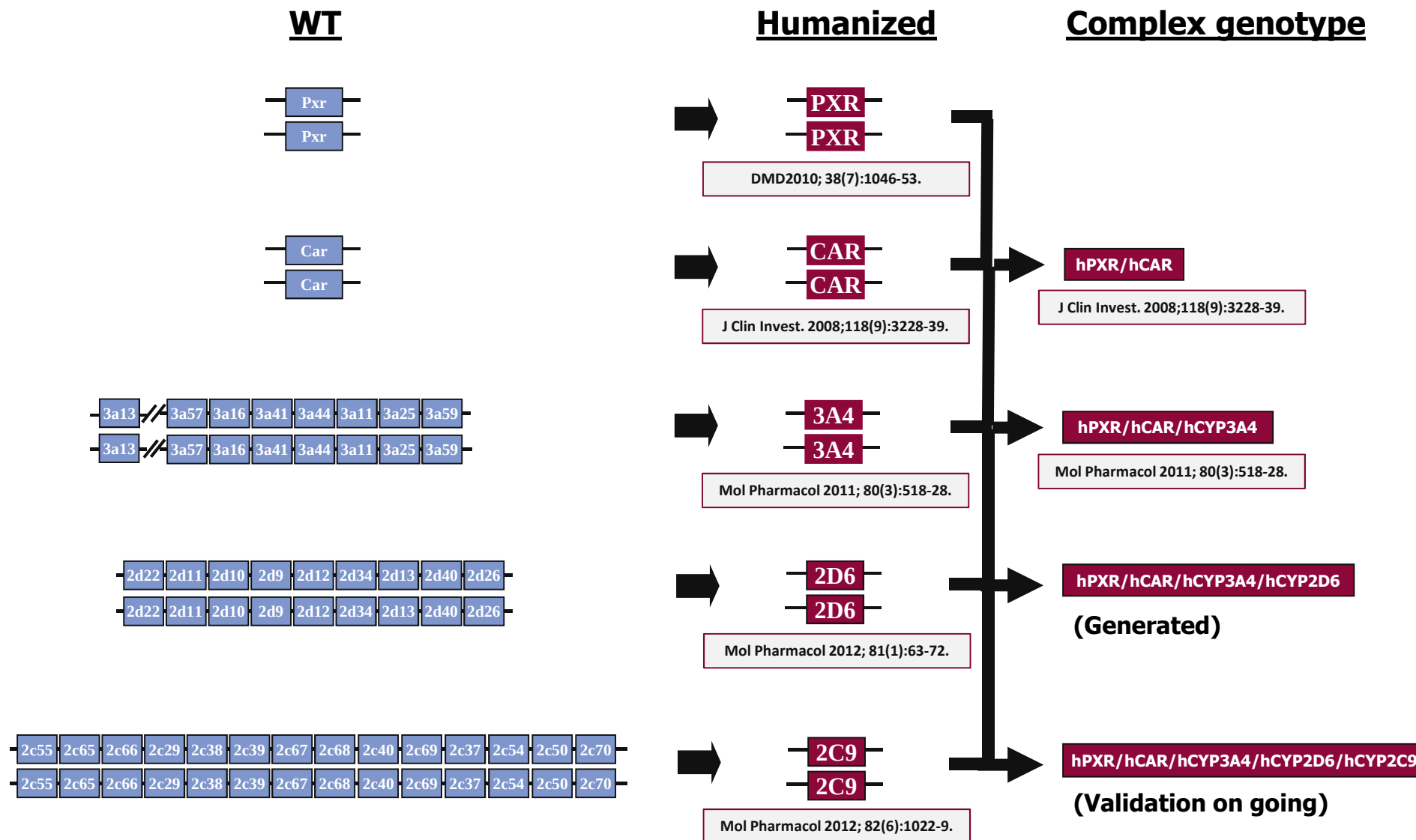


Xenoreceptor Panel		Cytochrome P450 Panel		Transporter Panel	
Knockouts	Humanized	Knockouts	Humanized	Knockouts	Humanized
Ahr	AHR	Cyp1a1/1a2	CYP1A1/1A2	Bcrp	BCRP*
Car	CAR	Cyp2c	CYP2C9	Mdr1a, Mdr1a/1b	MDR1*
Ppara α		Cyp2d	CYP2D6	Bcrp/Mdr1a/1b	
Pxr	PXR	Cyp3a (BL/6)	CYP3A4/3A7	Mrp1	
Pxr/Car	PXR/CAR	Cyp3a (FVB)	Liver CYP3A4	Mrp2	MRP2
Pxr/Car/Ahr	PXR/CAR/AHR		Gut CYP3A4	Oatp1a/1b	OATP1B1
Pxr/Car/Ppara α	PXR/CAR/PPAR α		Liver/Gut CYP3A4		OATP1B3
Pxr/Car/Ahr/Ppara α	PXR/CAR/AHR/PPAR α	Cyp2c/Cyp2d/Cyp3a			OATP1B1/1B3
				Oct1/2	
				Oat1/3	OAT1/3

Phase 2 Panel	Composite models		Transplanted Liver Humanized Mice
Humanized	Humanized		
UGT1A1	PXR/CAR/CYP3A4/3A7		FRG™ (in Collaboration with Yecuris Inc.)
	PXR/CAR/CYP3A4/3A7/CYP2D6		
	PXR/CAR/CYP3A4/3A7/CYP2D6/CYP2C9		

*coming soon

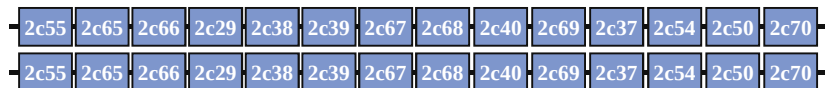
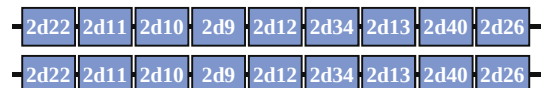
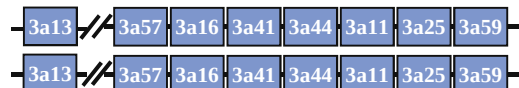
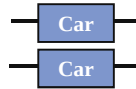
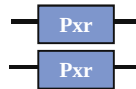
Towards a multiple humanized ADMET mouse model



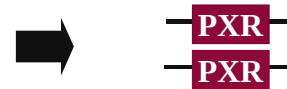
Use of hPXR/hCAR/hCYP3A4 mice for induction studies



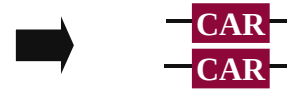
WT



Humanized



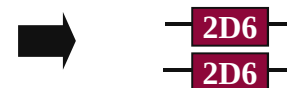
DMD2010; 38(7):1046-53.



J Clin Invest. 2008;118(9):3228-39.



Mol Pharmacol 2011; 80(3):518-28.



Mol Pharmacol 2012; 81(1):63-72.



Mol Pharmacol 2012; 82(6):1022-9.

Complex genotype

hPXR/hCAR

J Clin Invest. 2008;118(9):3228-39.

hPXR/hCAR/hCYP3A4

Mol Pharmacol 2011; 80(3):518-28.

hPXR/hCAR/hCYP3A4/hCYP2D6

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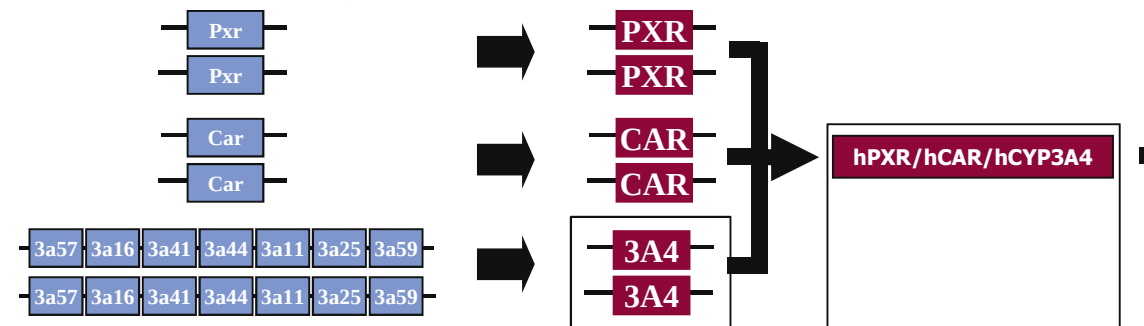
hPXR/hCAR/hCYP3A4/hCYP2D6/hCYP2C9

(First mice born)

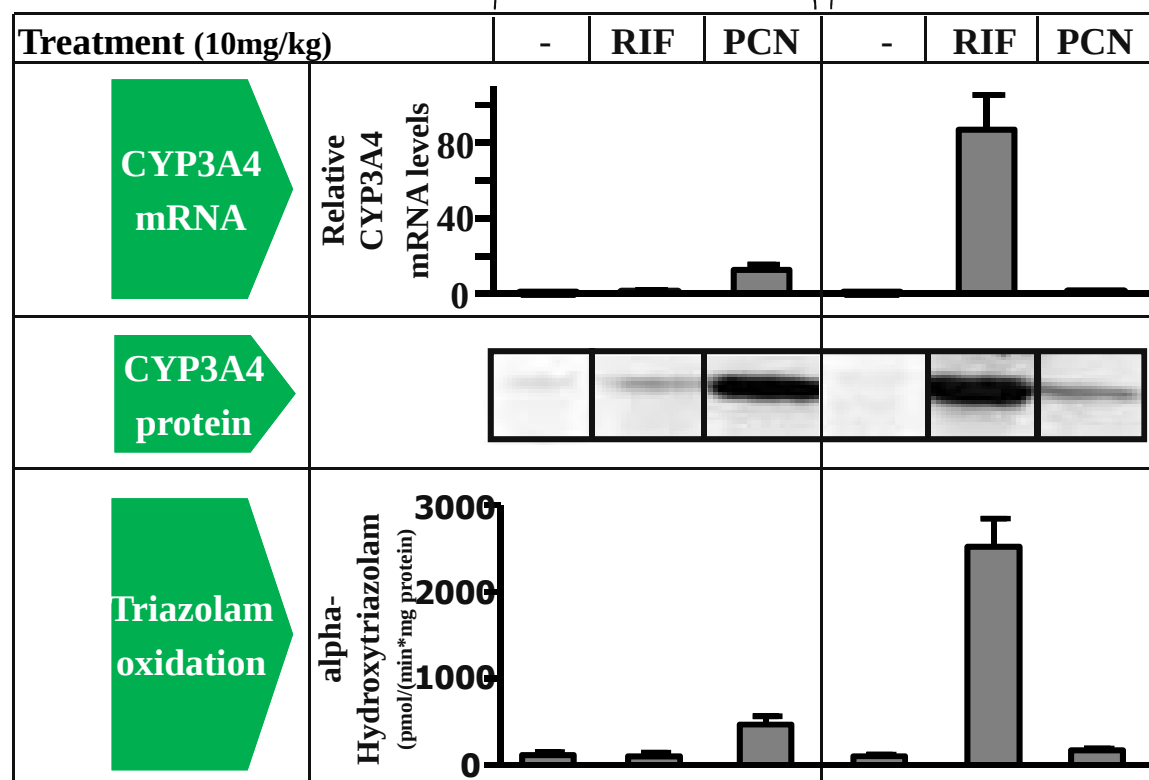
DDI prediction with triple humanized PXR/CAR/CYP3A4 mice

Quantitative prediction of human pregnane X receptor and cytochrome P450 3A4 mediated drug-drug interaction in a novel multiple humanized mouse line.

Hasegawa, Kapelyukh, Tahara, Seibler, Rode, Krueger, Lee, Wolf, Scheer; Mol Pharmacol. 2011 80(3):518-28.



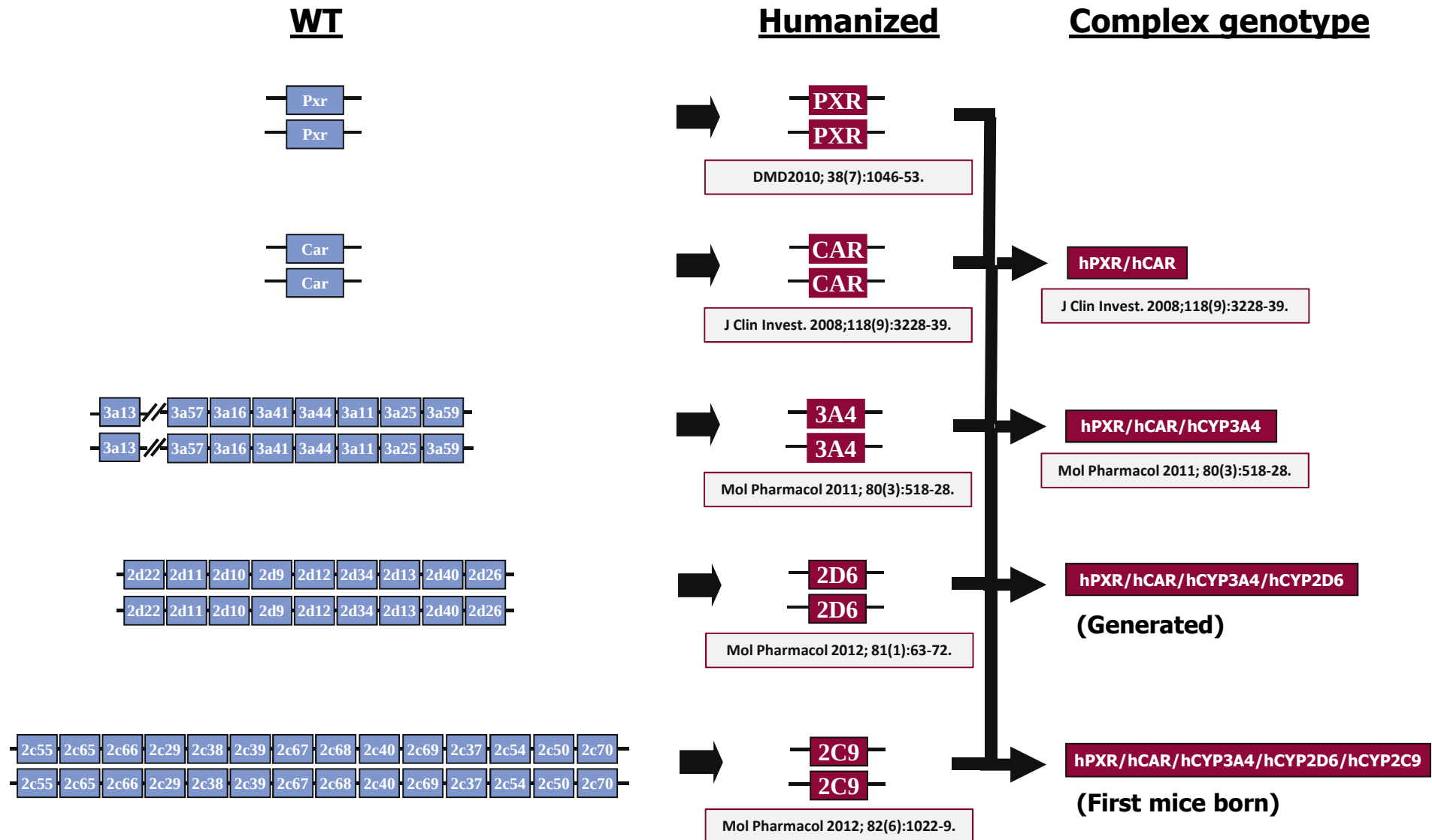
Prediction of clinical DDIs



human			
mouse*			
	Rifampicin	Sulfonpyrazole	Pioglitazone
	strong	medium	weak
Hepatic CYP3A4 induction	15-44-fold	4-10-fold	0-4-fold
AUC decrease of a co-administered CYP3A4 substrate	70-95%	38%	0-26%
	63-90%	15-37%	2%

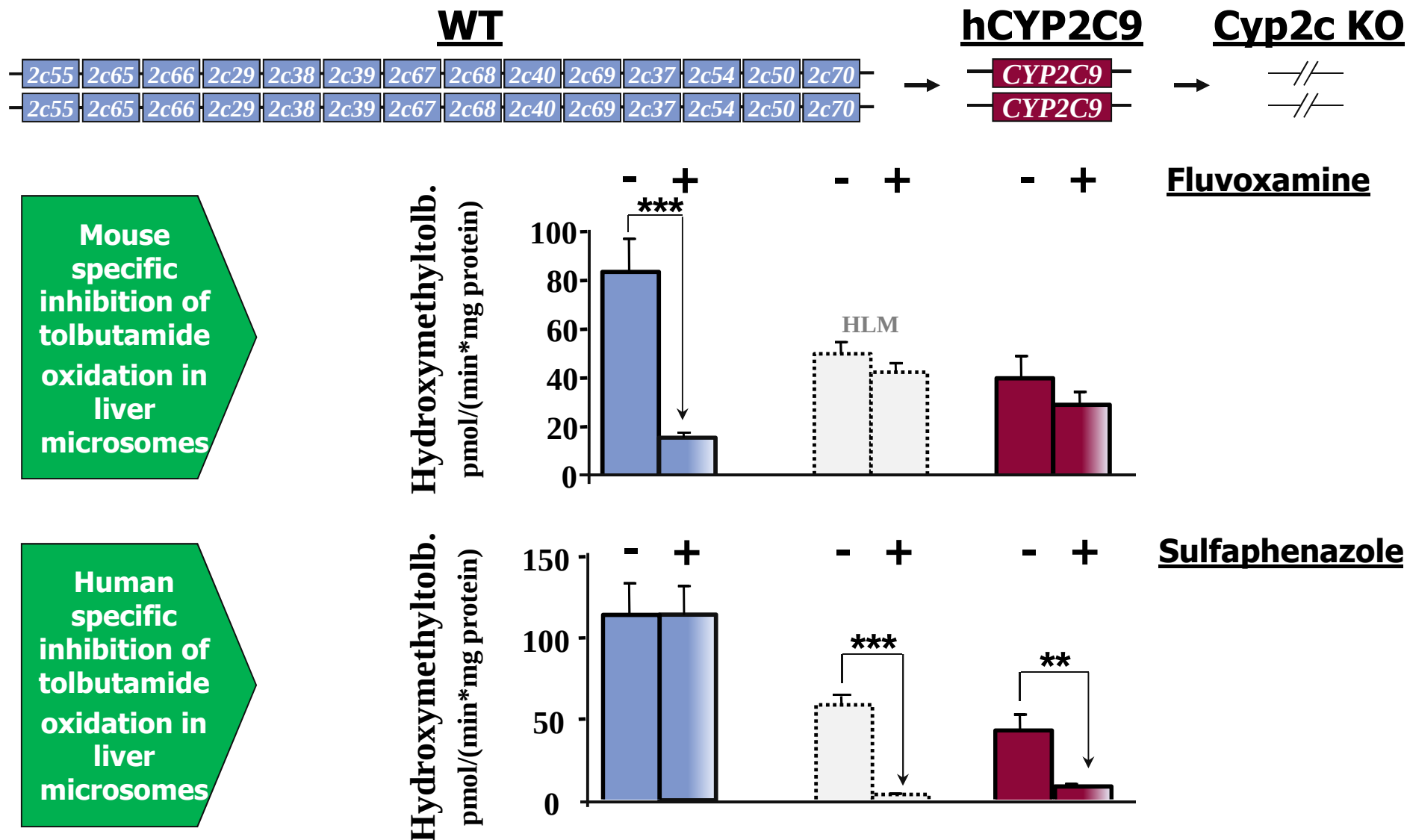
*All mouse data at clinically relevant doses

Use of cytochrome P450 humanized mice for inhibition studies



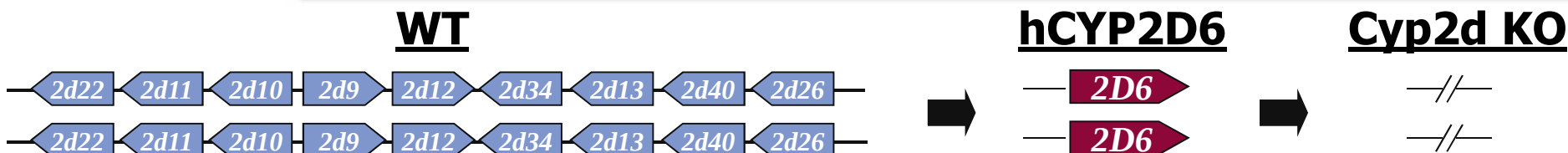
Inhibition studies in CYP2C9 humanized mice

Generation and characterization of novel
Cytochrome P450 Cyp2c gene cluster
knockout and CYP2C9 humanized mouse lines.
Scheer, Kapelyukh, Chatham, Rode, Buechel,
Wolf. Mol Pharmacol 2012; 82(6):1022-9.



Inhibition studies in CYP2D6 humanized mice

Modeling human cytochrome P450 2D6 metabolism and drug-drug interaction by a novel panel of knockout and humanized mouse lines.
Scheer, Kapelyukh, McEwan, Beuger, Stanley, Rode, Wolf. Mol Pharmacol. 2012;81(1):63-72.



Human specific inhibition of bufuralol oxidation in liver microsomes

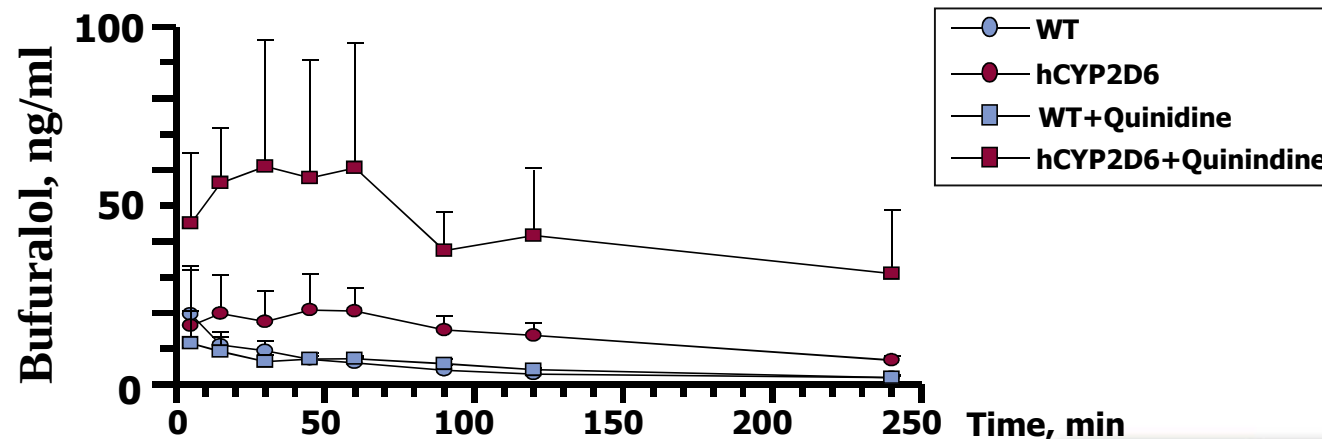
Hydroxybufuralol (pmol/min*mg protein)

2000
1000
0

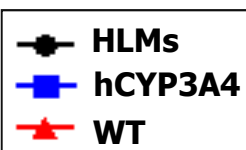


Quinidine (5μM)

Human specific inhibition of bufuralol oxidation in vivo



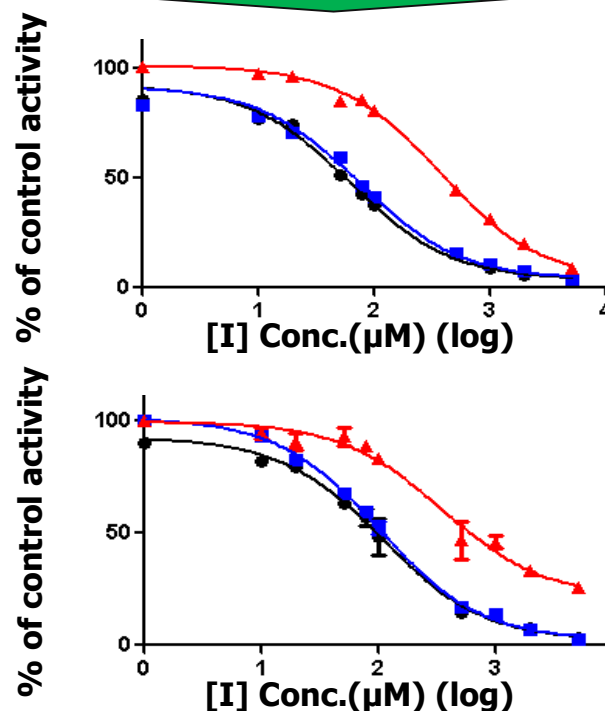
Inhibition of triazolam metabolism in CYP3A4 humanized mice (Liver microsomes)



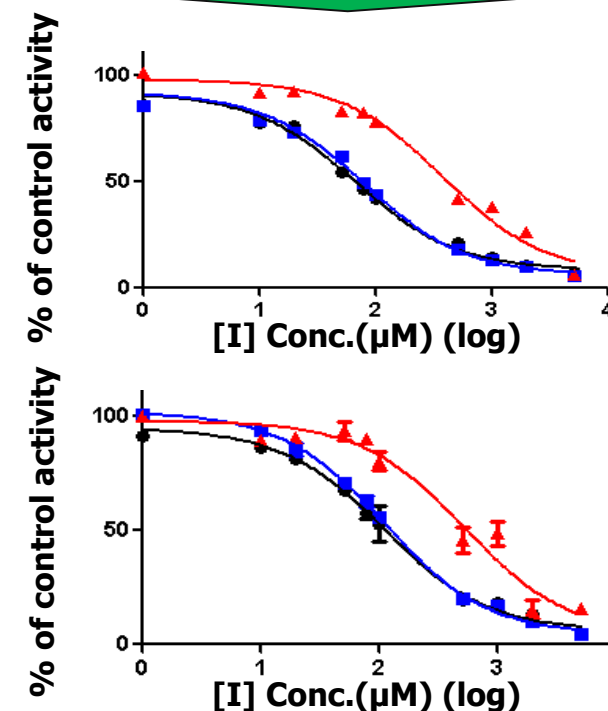
Erythromycin

Clarithromycin

1-OH triazolam formation



4-OH triazolam formation



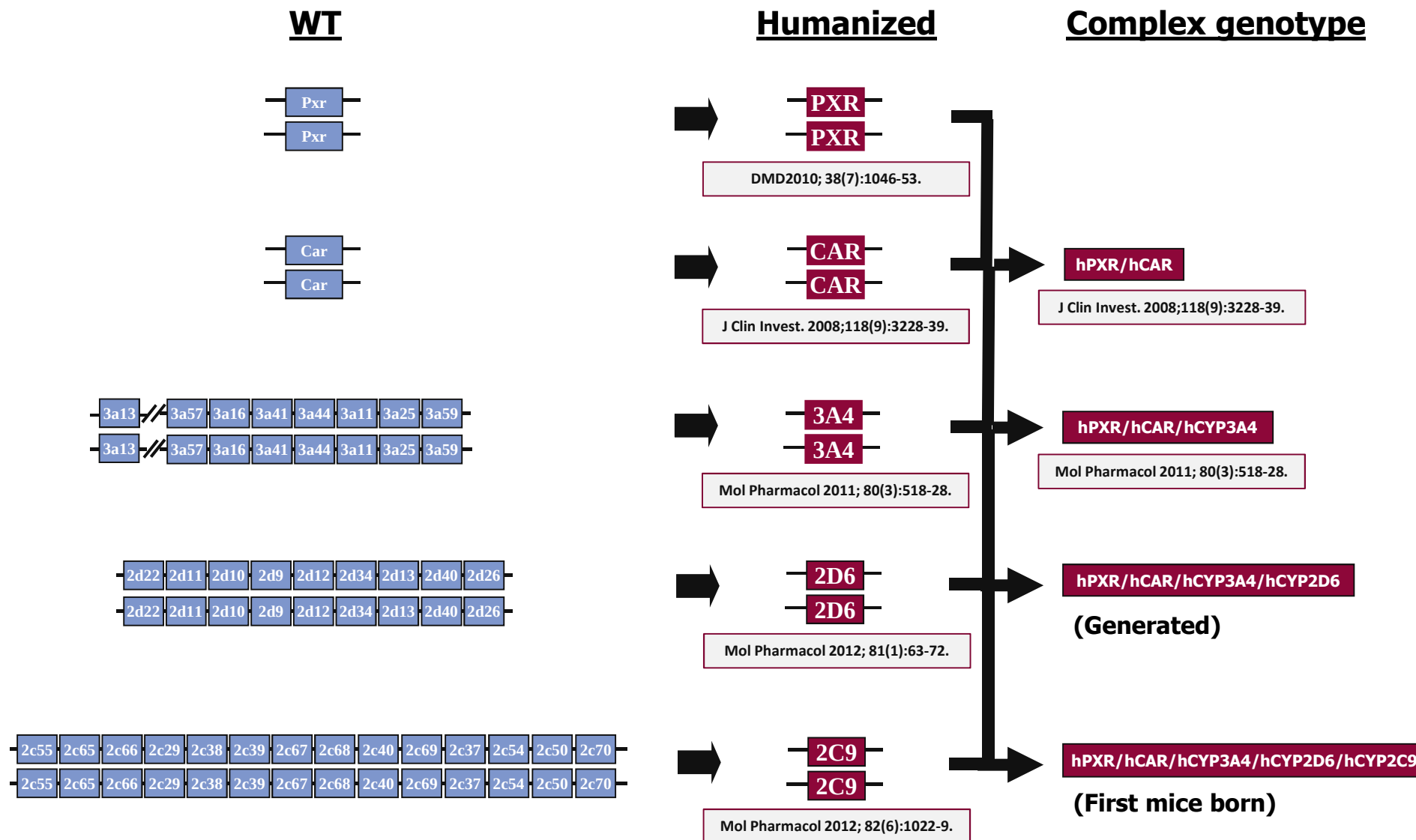
	1-OH Triazolam			4-OH Triazolam		
	HLM	hCYP3A4	WT	HLM	hCYP3A4	WT
Erythromycin IC ₅₀	62.0 µM	74.5 µM	725.4 µM	66.6 µM	80.0 µM	716.0 µM
Clarithromycin IC ₅₀	108.0 µM	101.9 µM	349.7 µM	111.5 µM	111.1 µM	527.7 µM



Bristol-Myers Squibb

Taconic

Use of cytochrome P450 humanized mice for human metabolite testing



Decision tree

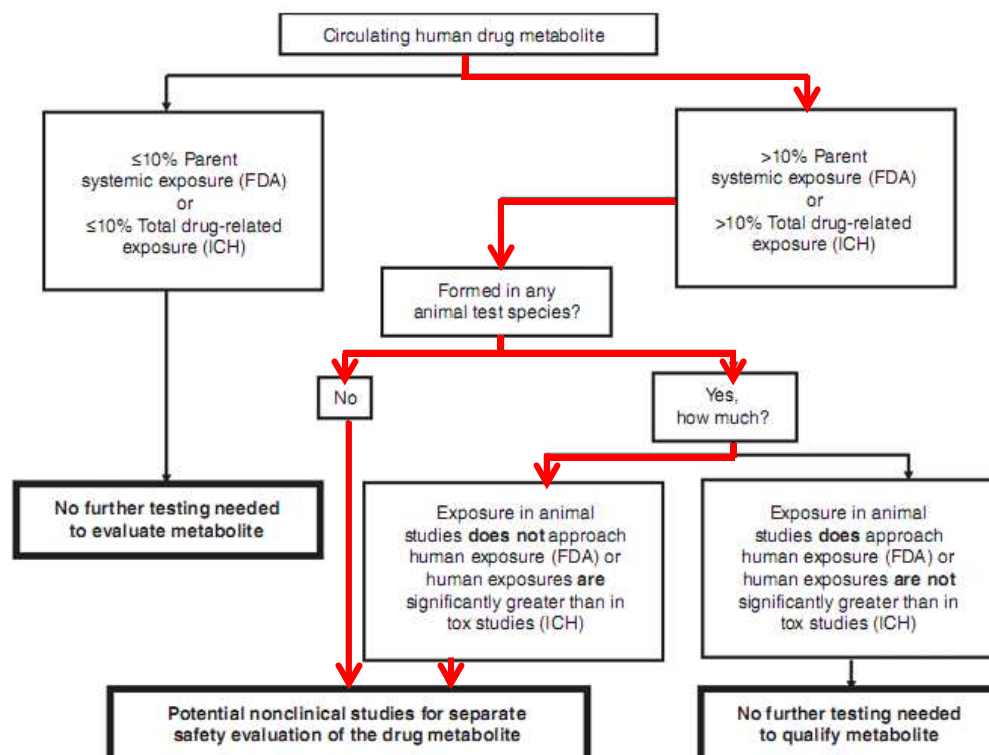


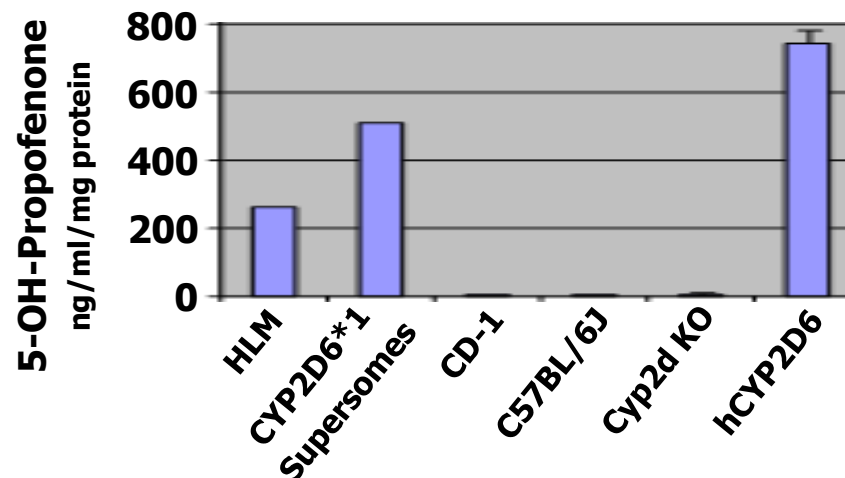
Figure 1 Combined decision tree for the implementation of the 2008 FDA and 2009 ICH guidances on the safety assessment of metabolites of drug candidates. The differences in the regulatory guidances on the decision-making criteria have not been formally resolved, although the criteria provided in the ICH guidance generally take precedence. FDA, US Food and Drug Administration; ICH, International Conference on Harmonisation.

Formation of unique and disproportionate metabolites in humanized CYP2D6 mice

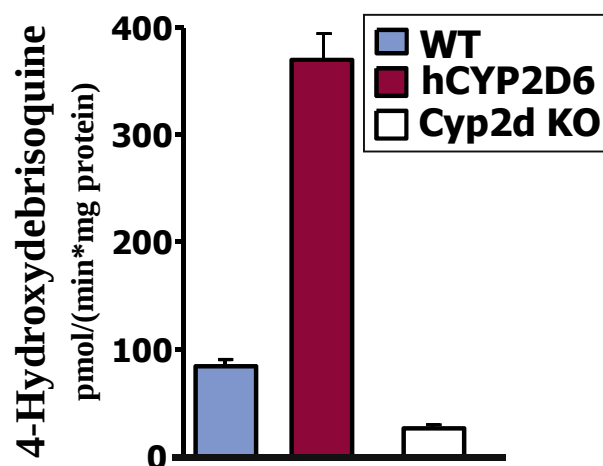
Modeling human cytochrome P450 2D6 metabolism and drug-drug interaction by a novel panel of knockout and humanized mouse lines.

Scheer, Kapelyukh, McEwan, Beuger, Stanley, Rode, Wolf. Mol Pharmacol. 2012;81(1):63-72.

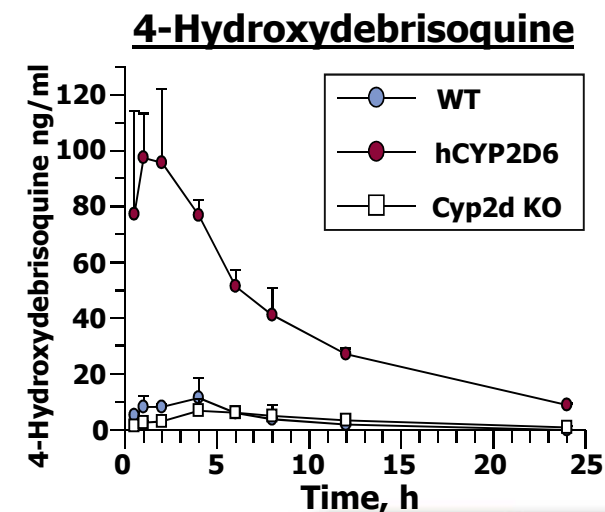
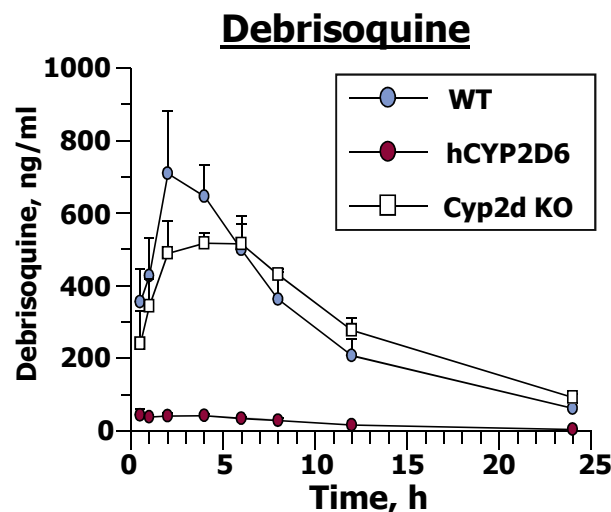
Propofenone oxidation by liver microsomes:



Debrisoquine oxidation by liver microsomes:



Pharmacokinetic of Debrisoquine & 4-Hydroxydebrisoquine:



A Panel of Translational Xenoreceptor, DME, Transporter & Liver Humanized Mouse Models



Xenoreceptor Panel		Cytochrome P450 Panel		Transporter Panel	
Knockouts	Humanized	Knockouts	Humanized	Knockouts	Humanized
Ahr	AHR	Cyp1a1/1a2	CYP1A1/1A2	Bcrp	BCRP*
Car	CAR	Cyp2c	CYP2C9	Mdr1a, Mdr1a/1b	MDR1*
Ppara α		Cyp2d	CYP2D6	Bcrp/Mdr1a/1b	
Pxr	PXR	Cyp3a (BL/6)	CYP3A4/3A7	Mrp1	
Pxr/Car	PXR/CAR	Cyp3a (FVB)	Liver CYP3A4	Mrp2	MRP2
Pxr/Car/Ahr	PXR/CAR/AHR		Gut CYP3A4	Oatp1a/1b	OATP1B1
Pxr/Car/Ppara α	PXR/CAR/PPAR α		Liver/Gut CYP3A4		OATP1B3
Pxr/Car/Ahr/Ppara α	PXR/CAR/AHR/PPAR α	Cyp2c/Cyp2d/Cyp3a			OATP1B1/1B3
				Oct1/2	
				Oat1/3	OAT1/3

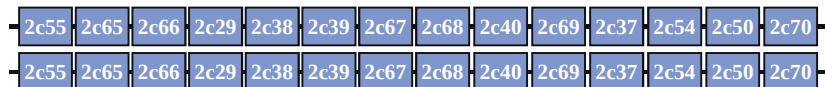
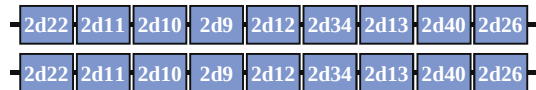
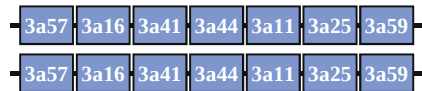
Phase 2 Panel	Composite models	Transplanted Liver Humanized Mice
Humanized	Humanized	
UGT1A1	PXR/CAR/CYP3A4/3A7	FRG™ (in Collaboration with Yecuris Inc.)
	PXR/CAR/CYP3A4/3A7/CYP2D6	
	PXR/CAR/CYP3A4/3A7/CYP2D6/CYP2C9	

*coming soon

Cyp3a/Cyp2c/Cyp2d KO mice



WT



Knockout



Mol Pharmacol 2011; 80(3):518-28.



Mol Pharmacol 2012; 81(1):63-72.

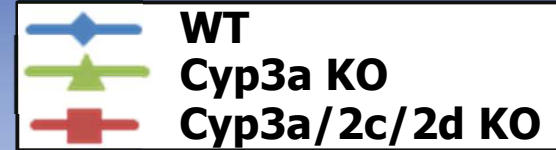


Mol Pharmacol 2012; 82(6):1022-9.

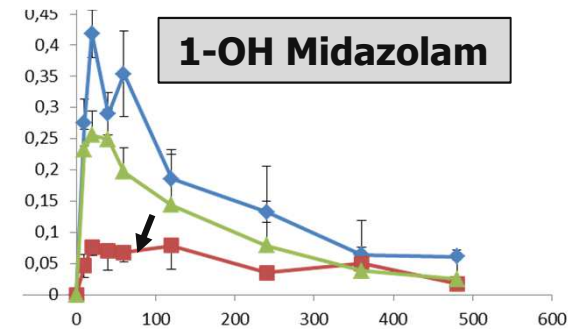
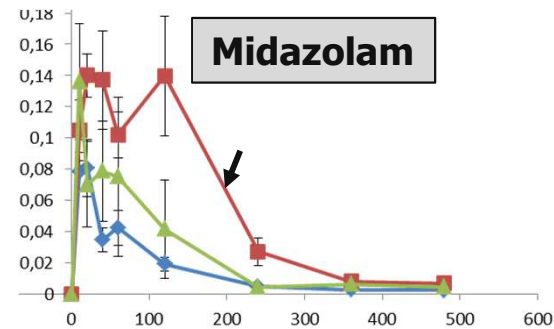
Complex genotype

Cyp3a/Cyp2c/Cyp2d KO

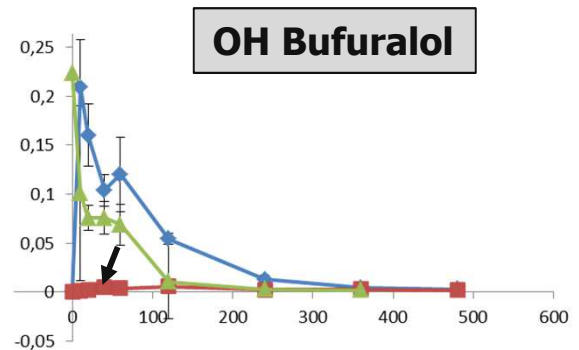
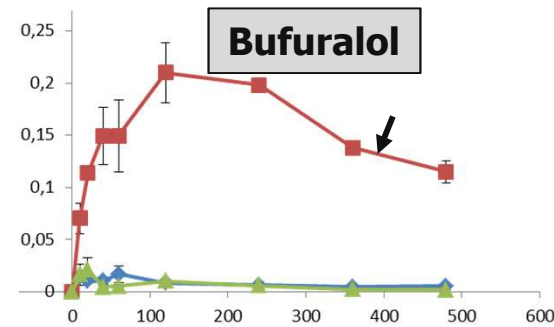
Cyp3a/Cyp2c/Cyp2d KO mice



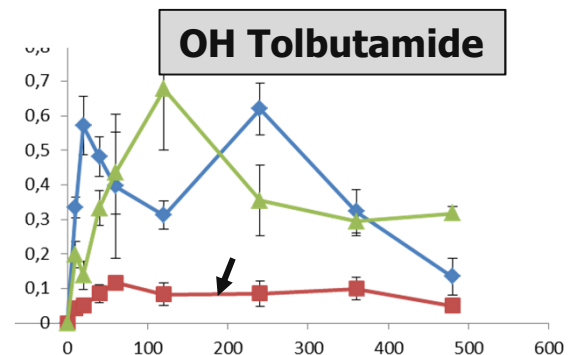
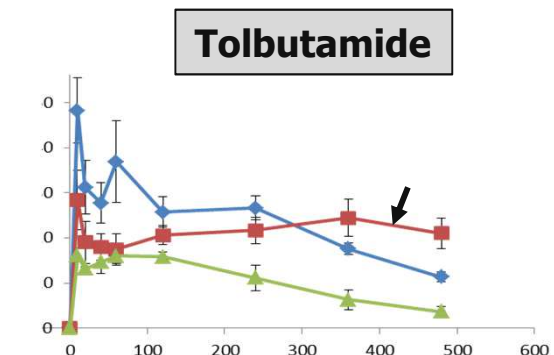
**CYP3A
substrate**



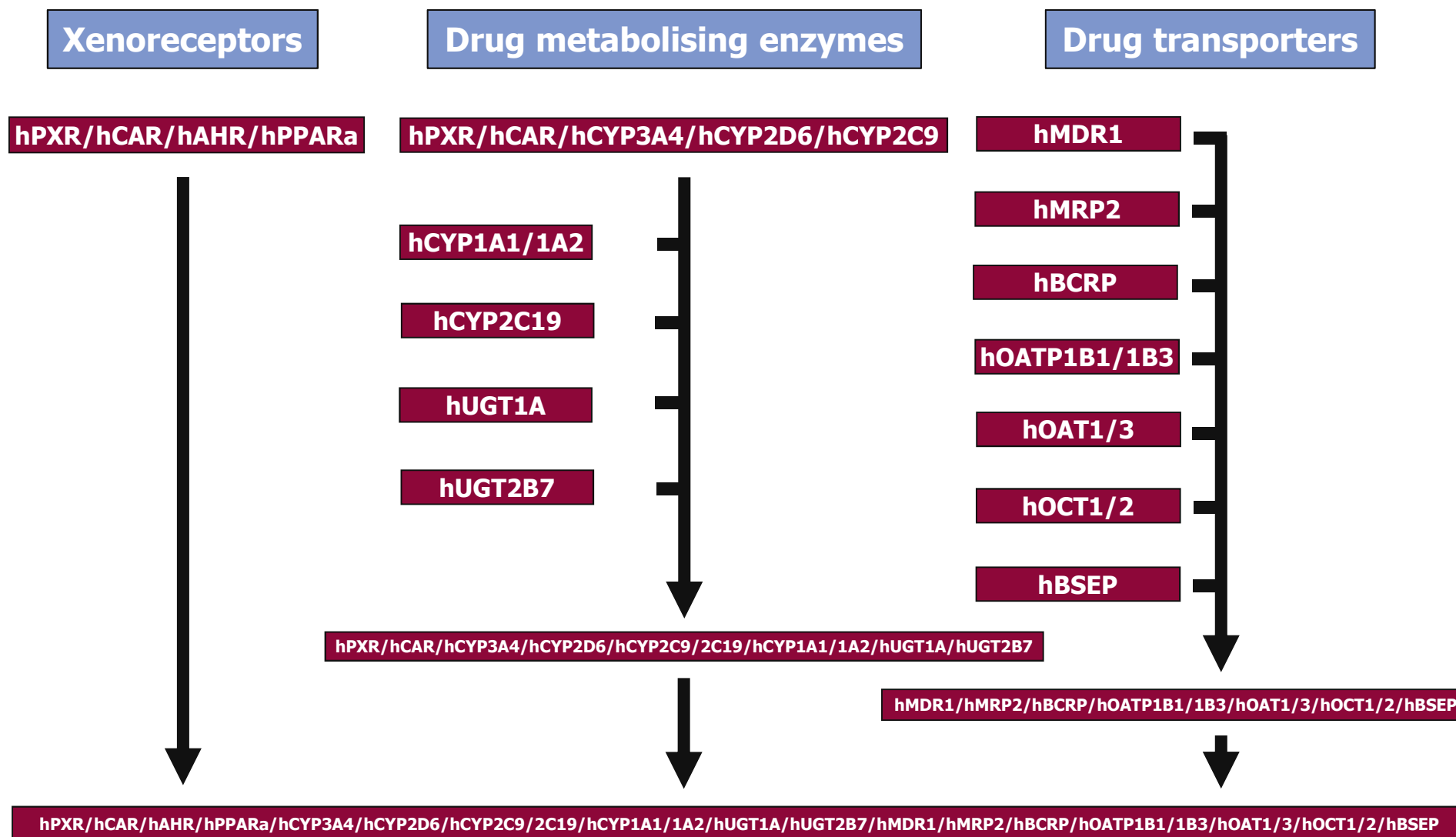
**CYP2D
substrate**



**CYP2C
substrate**



Future vision for genetically humanized mice



A Panel of Translational Xenoreceptor, DME, Transporter & Liver Humanized Mouse Models



Xenoreceptor Panel		Cytochrome P450 Panel		Transporter Panel	
Knockouts	Humanized	Knockouts	Humanized	Knockouts	Humanized
Ahr	AHR	Cyp1a1/1a2	CYP1A1/1A2	Bcrp	BCRP*
Car	CAR	Cyp2c	CYP2C9	Mdr1a, Mdr1a/1b	MDR1*
Ppara α		Cyp2d	CYP2D6	Bcrp/Mdr1a/1b	
Pxr	PXR	Cyp3a (BL/6)	CYP3A4/3A7	Mrp1	
Pxr/Car	PXR/CAR	Cyp3a (FVB)	Liver CYP3A4	Mrp2	MRP2
Pxr/Car/Ahr	PXR/CAR/AHR		Gut CYP3A4	Oatp1a/1b	OATP1B1
Pxr/Car/Ppara α	PXR/CAR/PPAR α		Liver/Gut CYP3A4		OATP1B3
Pxr/Car/Ahr/Ppara α	PXR/CAR/AHR/PPAR α	Cyp2c/Cyp2d/Cyp3a			OATP1B1/1B3
				Oct1/2	
				Oat1/3	OAT1/3

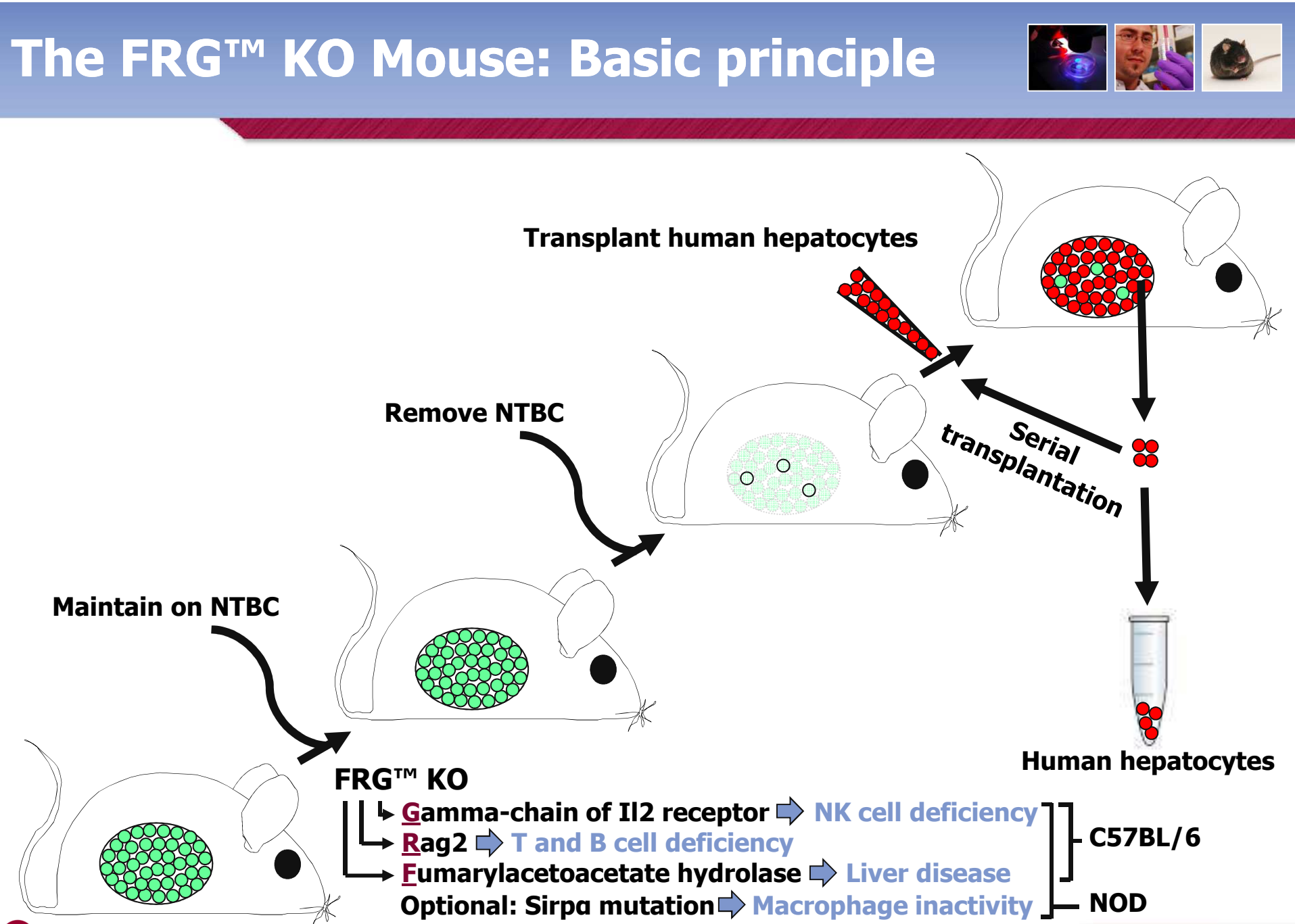
Phase 2 Panel	Composite models
Humanized	Humanized
UGT1A1	PXR/CAR/CYP3A4/3A7
	PXR/CAR/CYP3A4/3A7/CYP2D6
	PXR/CAR/CYP3A4/3A7/CYP2D6/CYP2C9

Transplanted Liver Humanized Mice

FRG™ (in Collaboration with Yecuris Inc.)

*coming soon

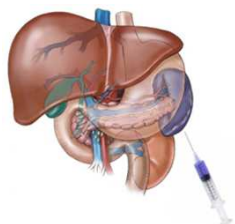
The FRG™ KO Mouse: Basic principle



Liver repopulation in FRG™ KO mice



Engraftment (.5-1M Cells)

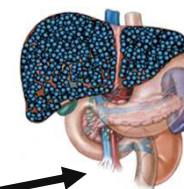


8 weeks (5-10M Cells)

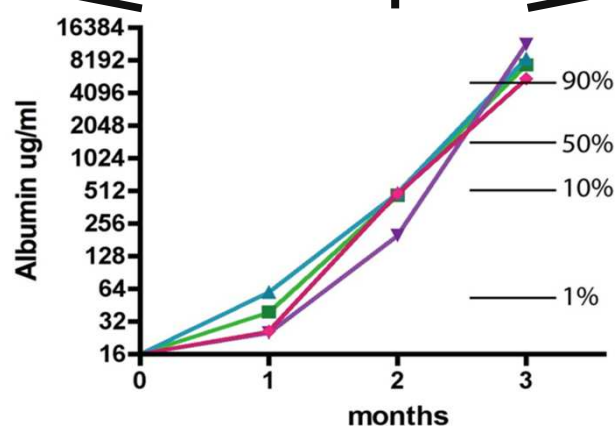


**200-500 µg/mL HAS
(1-5% repopulation)**

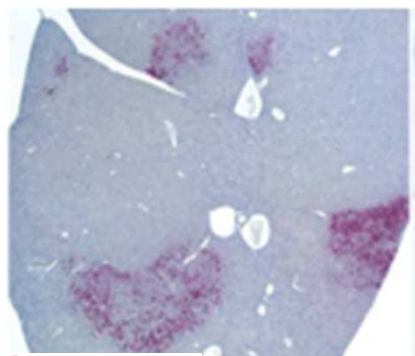
>12 weeks (80-150M Cells)



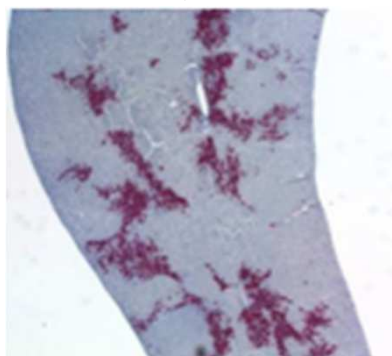
**2000-5000 µg/mL HAS
(50-98% repopulation)**



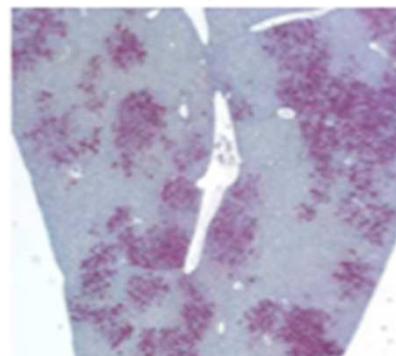
10-20%



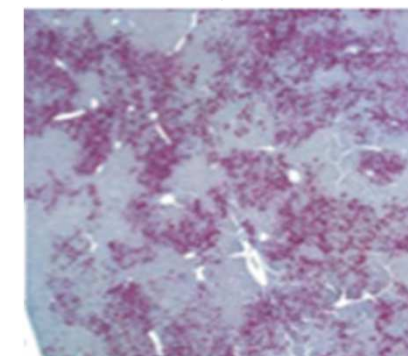
20-30%



30-50%



70-80%



Comparison of genetically and tissue humanized mouse models



Liver transplanted mice

Pros

- ✓ Humanization of all genes in given liver cell
- ✓ Only one mouse line required
- ✓ Ease of generating donor variability
- ✓ Allows for extraction of human hepatocytes
- ✓ Useful to study infectious liver diseases

Cons

- × Model has to be recreated continuously
- × Varying degree of humanization
- × Humanization of liver only
- × Presence of residual mouse hepatocytes
- × Higher unit price

Transgenic mice

Cons

- × Humanization of selected genes only
- × Requirement of various mouse lines
- × Effort of generating donor variability
- × No possibility to extract human tissues
- × No infection with human specific pathogens

Pros

- ✓ Permanent models without recreation
- ✓ Invariable quality of mice from each line
- ✓ Expression of human genes in other organs
- ✓ Human genes expressed in all liver cell
- ✓ Lower unit price

Prospects & limitations



Prospects

The described models may be valuable tools to predict certain aspects of drug responses in man, such as

- ◆ Drug-drug interactions
- ◆ Effects of human metabolites
- ◆ Bioavailability & clearance
- ◆ PKPD relation
- ◆ Drug-induced toxicity

Combination of genetically humanized mice into multiple humanized genotypes will further increase their utility.

Limitations

- ◆ Humanization restricted to selected genes (genetically humanized mice) or liver (transplanted models)
- ◆ General physiological differences between mice and humans
- ◆ Lower throughput and higher cost compared to in vitro studies
- ◆ Limited historical data

These *in vivo* technologies are suited to more detailed investigative studies & to select the most promising drug candidates before first test in man.

Acknowledgements



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(Validation of selected models)

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(Human metabolite studies)

UCSD:

Robert Tukey (UGT1A humanized mice)

Netherlands Cancer Institute:

Alfred Schinkel
(Transporter KOs, hOATP1B1/1B3 & hCYP3A4^{ApoE} & Villin)

Kyowa Kirin:

Harunobu Tahara, Maki Hasegawa
(hPXR/hCAR/hCYP3A4 DDI study)

BMS

Kamelia Behnia, Anne Rose
(CYP3A4 inhibition studies)

Yecuris

John Bial, Markus Grompe (FRG™ technology)

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